

## ANALYSIS OF VOLATILES IN AQUEOUS AND BIOLOGICAL SPECIMENS BY HEADSPACE GAS CHROMATOGRAPHY

### 10.1 POLICY

This test method may be used to confirm the presence of ethanol, acetone, isopropanol and methanol in aqueous and biological samples. Reporting of results following the application of this method will be contingent upon a thorough review and acceptance of quality control data and the qualification of individual results under the criteria for acceptance.

Any adjustments or deviations from the procedures below must be approved by a member of TLD Management, and appropriately documented in the batch file.

### 10.2 PURPOSE

The purpose of this technical procedure is to describe the identification and/or quantitation of ethanol, acetone, isopropanol (2-propanol) and methanol in aqueous and biological samples by headspace gas chromatography (HSGC) using an alcohol analysis capillary column and a flame ionization detector (FID). This procedure will serve as the laboratory document describing sample preparation, instrumental analysis, data analysis, and criteria for acceptance of volatile compounds.

### 10.3 PRINCIPLE

There is a direct relationship between the concentration of a volatile substance (e.g. ethanol) dissolved in a liquid (e.g., blood) and the concentration of the volatile substance in the vapor above the solution (headspace) for a given temperature, based on Henry's Law.

An aqueous or biological specimen is measured into a vial and then diluted with a measured volume of internal standard (n-propanol, 1-propanol). The vial is then sealed with a septum-equipped airtight seal. After a short incubation period at 70°C, the vial is pressurized and a measured aliquot of the headspace is transferred to the gas chromatograph for analysis.

Ethanol is resolved from other volatiles, such as acetone, isopropanol and methanol. Identification is by comparison of retention times of observed analytes to those present in the calibrators. Quantitation is accomplished by multilevel calibration. Each calibration level corresponds to a calibration sample with a known concentration of components. Confirmation is performed on a separate, identical instrument that is equipped with a separation column having different selectivity.

### 10.4 SPECIMENS

10.4.1 The specimen volume is 0.2 mL.

10.4.2 Specimens include whole blood, serum, plasma, urine, vitreous humor, tissue homogenate and aqueous samples. [NOTE: For Liquor and Cannabis Board sample testing, refer to the *Policy on Testing and Reporting Results for Liquor and Cannabis Board Samples (P48-3)*.]

10.4.3 Dilutions of specimens may be analyzed at the Forensic Scientist's discretion; in addition, the specimen may be analyzed at standard volume, as dictated by screening results, to ensure that concentrations of all volatiles present are within the reporting range of the test method. The volume of n-propanol internal

standard added is 2 mL for standard volume specimens and dilutions of specimens (adjustment of diluted specimens to standard volume is not required).

10.4.4 Analysis of larger specimen volumes must be approved and documented.

NOTE: Ethanol and sevoflurane co-elute on the J&W DBALC2 capillary column. For specimens containing both ethanol and sevoflurane, ethanol confirmation must be performed on the J&W DBALC1 column only.

10.4.5 If volatiles other than ethanol, acetone, isopropanol or methanol are suspected in a specimen, two additional samples of the specimen are tested in order to identify/confirm the presence of the volatile compound (for analysis by HSGC and gas chromatography – mass spectrometry). See *Identification/ Confirmation of Miscellaneous Volatiles in Aqueous and Biological Specimens by Headspace Gas Chromatography and Gas Chromatography – Mass Spectrometry (TCmv12726)* for miscellaneous volatile testing.

## 10.5 EQUIPMENT AND MATERIALS

### 10.5.1 EQUIPMENT

- 10.5.1.1 Calibrated, adjustable piston pipettes with disposable tips
- 10.5.1.2 Disposable transfer pipettes (glass or polyethylene)
- 10.5.1.3 Microlab® 600 Autopipette, Hamilton Automatic Diluter, or equivalent
- 10.5.1.4 Headspace autosampler vials (10 mL) and crimp tops
- 10.5.1.5 Cap crimper
- 10.5.1.6 Agilent (Hewlett Packard) 7694/G1888 headspace autosampler, or equivalent
- 10.5.1.7 Agilent (Hewlett Packard) 6890 gas chromatograph equipped with either a J&W DBALC1 capillary column (30 m x 0.53 mm ID x 3 µm film thickness) or a J&W DBALC2 capillary column (30 m x 0.53 mm ID x 2 µm film thickness), or equivalent
- 10.5.1.8 Computer system equipped with Agilent (Hewlett-Packard) ChemStation or OpenLab software

### 10.5.2 MATERIALS

- 10.5.2.1 Deionized water, laboratory grade (DI H<sub>2</sub>O)
- 10.5.2.2 Disposable 12 x 75 mm tubes with closures

## 10.6 STANDARDS, CALIBRATORS AND CONTROLS

### 10.6.1 STANDARDS

- 10.6.1.1 Internal standard (n-propanol) is prepared and verified according to the *Procedure for the Verification of n-Propanol Internal Standard (PTis12501)*.

### 10.6.2 CALIBRATORS

- 10.6.2.1 Ethanol calibrators are prepared and verified according to the *Procedure for the Verification of Ethanol Calibrators (PTec12500)*.
- 10.6.2.2 Mixed volatile calibrators are purchased from an approved certified reference material (CRM) provider, and verified according to the instructions on the *Combined Mixed Volatile Verification Worksheet (CVVerWS)*. Verification is required prior to use.

NOTE: The multicomponent alcohol CRM materials contain ethanol. The presence of ethanol in these materials is for demonstration of chromatographic separation from other volatile compounds only. Batch acceptability and reporting of ethanol results are determined from the ethanol-only calibration data. Ethanol results from data processed using the mixed volatile method (from three-point mixed volatile calibration) have not been validated, and are not reported in specimen data.

### 10.6.3 CONTROLS

- 10.6.3.1 Commercially prepared ethanol controls (CTRL) are purchased for use with each assay. The source and lot number of each control is documented in the *Alcohol Control Log (TLD\_AlcCtl\_Log)*. The controls are verified according to the instructions on the *Combined Ethanol Verification Worksheet (CEVerWS)*. Verification is required prior to use. Controls are stored per manufacturer specifications.

- a. Four ethanol-only controls are used, at the following concentrations:

CTRL 1	0.04 CTRL	0.04 g/100 mL
CTRL 2	0.10 CTRL	0.10 g/100 mL
CTRL 3	0.20 CTRL	0.20 g/100 mL
CTRL 4	0.40 CTRL	0.40 g/100 mL

- b. Controls other than the aforementioned may be approved for use by the State Toxicologist or QA Manager, with appropriate documentation.

- c. Ethanol controls are verified and considered approved for use when quantifying within the following inclusive ranges:

CTRL 1	0.038 – 0.042 g/100 mL
CTRL 2	0.095 – 0.105 g/100 mL
CTRL 3	0.190 – 0.210 g/100 mL
CTRL 4	0.380 – 0.420 g/100 mL

[NOTE: These ranges apply for initial verification of controls only.]

- 10.6.3.2 For initial screening of specimens for mixed volatile compounds (acetone, isopropanol and methanol), a mixed volatile positive screening control is included in the batch. The screening control is prepared and verified as described in the *Procedure for the Verification of Mixed Volatile Calibrators and Controls (PTvc12502)* and recorded on the *Combined Mixed Volatile Verification Worksheet (CVVerWS)*.

10.6.3.3 When quantifying a mixed volatile compound in the batch, a quantitative mixed volatile control is prepared as a dilution of a high concentration (4000 µg/mL) multicomponent alcohol CRM.

- a. The quantitative positive mixed volatile control is used at the following concentration:

CTRL 5          60 CTRL          60 mg/dL

- b. Controls other than the aforementioned may be approved for use by the State Toxicologist or QA Manager, with appropriate documentation.

See NOTE in 10.6.2.2.

## 10.7 SAMPLE PREPARATION

10.7.1 Label 10 mL headspace vials for each member of the test batch (blank, negative controls, calibrators, positive controls, specimen samples, etc.) The batch should be set up according to the following suggested sequence:

1. Blank (DI H <sub>2</sub> O, no ISTD)	12. CTRL 4 (0.40 g/100 mL)
2. CAL 1 (0.079 g/100 mL)	13. CTRL 5 (60 mg/dL) Volatile Control
3. CAL 2 (0.158 g/100 mL)	14. Negative Control
4. CAL 3 (0.316 g/100 mL)	15. Specimen #1
5. Negative Control (DI H <sub>2</sub> O, plus ISTD)	16. Specimen #2
6. VOL 1 (10 mg/dL)	17. Specimen #3
7. VOL 2 (50 mg/dL)	18. (etc.)
8. VOL 3 (100 mg/dL)	Insert a positive ethanol control & negative control after every 10 samples
9. CTRL 1 (0.04 g/100 mL)	Conclude the sequence with CTRL 5, positive ethanol control & negative control
10. CTRL 2 (0.10 g/100 mL)	Confirmation batches for specimens containing ethanol only will omit CTRL 5 at the end of the sequence
11. CTRL 3 (0.20 g/100 mL)	

[NOTE: It is advisable to run an extra blank following any badly decomposed specimen or where volatiles other than ethanol, acetone, isopropanol or methanol are expected (e.g., toluene, difluoroethane).]

For initial batch screening for mixed volatile compounds, the use of a single aliquot of the positive mixed volatile screening control is analyzed (following ethanol CTRL 1-4), omitting the mixed volatile CRM calibrators and CTRL 5. This injection of the mixed volatile screening control, and the conclusion of the sequence with a positive ethanol control and negative control, serves to bracket case specimens with positive controls.

Confirmation batches must be run on a different GC column than the initial screen and need only include mixed volatile calibrators 1-3 and CTRL 5 (bracketing case specimens) if a case in the batch requires quantitation of acetone, isopropanol and/or methanol.

10.7.2 Equilibrate specimens to room temperature and mix before opening under a biohazard hood. Blood specimens are inspected to ensure the blood is mobile. If necessary, the sample may be sonicated or homogenized.

- 10.7.3 If the batch includes quantitation of mixed volatile compounds, prepare the mixed volatile positive quantitative control (CTRL 5) solution.
- Using a calibrated pipette, add 150  $\mu\text{L}$  of the high concentration (4000  $\mu\text{g}/\text{mL}$ ) multicomponent alcohol CRM and 850  $\mu\text{L}$  DI  $\text{H}_2\text{O}$  to a clean, labeled 12 x 75mm tube. Cap the tube and briefly vortex mix.
  - The solution is for use on day of preparation only. The tube is labeled with the initials of the person preparing the solution, the identity and the concentration. The preparing analyst will record the preparation information on their batch worklist (pipette ID, lot number of multicomponent alcohol CRM used to prepare solution). Subsequent analysts using the prepared control solution will record the initials of the preparing analyst and the date prepared on their batch worklist or sequence.
- 10.7.4 Add approximately-2.2 mL DI  $\text{H}_2\text{O}$  into the vial labeled blank and seal the vial tightly.
- 10.7.5 Using the auto-pipetter, aliquot 200  $\mu\text{L}$  of the calibrators, controls or specimens and 2 mL of the internal standard solution into the respectively labeled headspace vial.
- 10.7.6 Seal the vial tightly.
- 10.7.7 Between each aliquot, rinse and wash the pipette tip appropriately (e.g., rinse pipette tip with diluted bleach and/or DI  $\text{H}_2\text{O}$ . Repeat if necessary.)

## 10.8 INSTRUMENTAL PARAMETERS

- 10.8.1 Load and edit a sequence on the HSGC. Enter the blank, calibrators, controls and specimens into the sequence table, and identify them appropriately under Sample Type.
- 10.8.2 Place each headspace vial in its respective position on the headspace autosampler and verify this placement against the sequence.
- 10.8.3 Run the sequence under method BLDALCO. [Note: The method name may contain a numeric suffix to differentiate between instruments; for example BLDALCO5 for headspace instrument 5.]
- 10.8.4 If the batch size is larger than the capacity of the headspace autosampler, the analyst may run the remaining specimens and controls on the same instrument after the initial sequence has completed.

The initial sequence must end with a positive and negative control. The appended sequence must begin and end with both a positive and negative control. Any appended sequence must be injected on the same day as the initial sequence and aliquoted at the same time as the entire batch.

## 10.9 REINJECTION

If necessary, reinjection of samples is performed after completion of the initial sequence. The reason(s) for reinjection (e.g., for specimens that follow a decomposed sample or one that contained difluoroethane) will be documented in the batch.

The original calibration for the batch must be valid in order to perform partial batch reinjection (described below). If the original calibration has been replaced, the calibration

for the batch must also be reinjected (vials 1-14 listed in 10.7.1), omitting 6-8 and 13 for ethanol only or 2-5 and 9-12 for mixed volatile only.

10.9.1 Should reinjection of a known sample (blank, calibrator, positive or negative control) be necessary, this may be done as a single injection.

10.9.2 Reinjections of unknown specimens must be bracketed by both positive and negative controls.

10.9.3 Examples:

- a. If the specimen(s) contains only ethanol, the reinjection sequence is as follows:

1. POS CTRL (ethanol)
2. Negative Control (DI H <sub>2</sub> O, plus ISTD)
3. Specimen (up to 10 injections)
4. POS CTRL (ethanol)
5. Negative Control (DI H <sub>2</sub> O, plus ISTD)

- b. If the specimen(s) contains only mixed volatiles, the reinjection sequence is as follows:

1. CTRL 5 (60 mg/dL) Volatile Control
2. Negative Control (DI H <sub>2</sub> O, plus ISTD)
3. Specimen (up to 10 injections)
4. CTRL 5 (60 mg/dL) Volatile Control
5. Negative Control (DI H <sub>2</sub> O, plus ISTD)

- c. If the specimen(s) contains ethanol and mixed volatiles, the reinjection sequence is as follows:

1. CTRL 5 (60 mg/dL) Volatile Control
2. POS CTRL (ethanol)
3. Negative Control (DI H <sub>2</sub> O, plus ISTD)
4. Specimen (up to 10 injections)
5. CTRL 5 (60 mg/dL) Volatile Control
6. POS CTRL (ethanol)
7. Negative Control (DI H <sub>2</sub> O, plus ISTD)

10.9.4 If the specimen is negative upon initial injection, the reinjection sequence in 10.9.3.a is used.

10.9.5 If the initial injection of the specimen has no results (e.g., due to clogged needle, loose cap), the reinjection sequence in 10.9.3.c is used. For a screening batch, replace CTRL 5 in vial 1 with the mixed volatile screening control and omit the CTRL 5 in vial 5.

10.9.6 Reinjected controls are subject to those evaluation criteria described in 10.11.3

## 10.10 DATA ANALYSIS

10.10.1 Analysis of the batch data is conducted using the instrumental data analysis software in ChemStation or OpenLab. Ethanol data is processed using the

BLDALCO method. Mixed volatile data is acquired using the BLDALCO method and processed using the mixed volatile quantitation method VOL (quantitative analysis) or the mixed volatile screening method MVSCREEN. The data analysis method name may contain a numeric suffix, as described in the note in 10.8.3.

10.10.2 Quantitative calculations are generated by internal standard, multi-point, linear regression with equal weighting and the origin included.

10.10.3 The assigned value for each ethanol calibrator level in the batch calibration table matches those for the listed lot numbers as verified against a NIST curve. Assigned values for each mixed volatile calibrator level are the nominal concentrations of the calibrators.

10.10.4 Printed reports for each vial in the batch are generated for review, along with a copy of the calibration table(s) applied to the batch.

10.10.5 Technical review of the batch is conducted according to the criteria listed below.

#### 10.11 CRITERIA FOR BATCH ACCEPTANCE

If the analysis of the batch meets the criteria listed below, the results for the specimens are accepted.

##### 10.11.1 Blank

10.11.1.1 The blank shall be devoid of any significant peaks<sup>1</sup>.

##### 10.11.2 Calibrators and calibration curves

###### 10.11.2.1 Ethanol

- a. Chromatographic peaks for ethanol and n-propanol shall appear symmetrical (i.e., no co-elution, split peaks, or shoulders).
- b. Retention times for ethanol and n-propanol shall be within  $\pm 2\%$  of those in ethanol calibrator 3. These are inclusive ranges.
- c. Quantitative results for ethanol in each calibrator shall be within  $\pm 10\%$  of their nominal values. Result comparisons will use values to three decimal places. These are inclusive ranges.
- d. The calibration curve for ethanol shall have a correlation coefficient  $\geq 0.99$ .

###### 10.11.2.2 Mixed Volatile

- a. Chromatographic peaks for acetone, isopropanol, methanol and n-propanol shall appear symmetrical (i.e., no co-elution, split peaks, or shoulders).
- b. Retention times for acetone, isopropanol, methanol and n-propanol shall be within  $\pm 2\%$  of those in mixed volatile calibrator 3 (for quantitation). These are inclusive ranges.

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<sup>1</sup> Peaks appearing in the blank, calibrators, or positive and negative controls that are fully resolved from any volatile compound or internal standard are considered extraneous and not significant.

- c. For quantitative confirmation, calculated results for acetone, isopropanol and methanol in each mixed volatile calibrator shall be within  $\pm 10\%$  of their nominal values. For calibrator 1 (target concentration 10 mg/dL), result comparisons will use whole integer values truncated after the first decimal place (acceptable range 9.0 – 11.0 mg/dL). For calibrators 2 and 3, result comparisons will use truncated, whole integer values in units of mg/dL. These are inclusive ranges.
- e. The calibration curves for acetone, isopropanol and methanol shall have correlation coefficients  $\geq 0.99$ .
- f. Evaluation of ethanol performance in the mixed volatile calibrators is not used in determining the acceptability of a batch for reporting of ethanol (see NOTE in 10.6.2.2).

### 10.11.3 Controls

#### 10.11.3.1 Negative controls

- a. The negative control(s) shall not identify ethanol at or above 0.005 g/100 mL. Identification is based on acceptable retention time matching and an integrated, symmetrical peak. All negative controls (ethanol) must meet these criteria for the batch to be accepted for reporting of ethanol.
- b. The negative control(s) shall not identify acetone, isopropanol or methanol at or above 10 mg/dL, based on criteria listed in 10.11.3.1.a above. All negative controls must meet these criteria for the batch to be accepted for reporting of a mixed volatile compound. Failure to meet criteria for one mixed volatile compound does not invalidate the acceptability of another mixed volatile compound.

#### 10.11.3.2 Positive ethanol controls

- a. Chromatographic peaks for ethanol and n-propanol shall appear symmetrical.
- b. Retention times for ethanol and n-propanol shall be within  $\pm 2\%$  of those in ethanol calibrator 3. These are inclusive ranges.
- c. Quantitative results for ethanol in each control shall be within  $\pm 7.5\%$  of their nominal values. Result comparisons will use values to three decimal places. These are inclusive ranges.
- d. All positive ethanol controls must meet these criteria for the batch to be accepted.

#### 10.11.3.3 Positive mixed volatile control

NOTE: For initial identification of acetone, isopropanol and methanol, the positive mixed volatile screening control must meet criteria in 10.11.3.3.a, below.

- a. Chromatographic peaks for acetone, isopropanol, methanol and n-propanol shall appear symmetrical.

- b. Retention times for acetone, isopropanol, methanol and n-propanol shall be within  $\pm 2\%$  of those in mixed volatile calibrator 3. These are inclusive ranges.
- c. Quantitative results for acetone, isopropanol and methanol in CTRL 5 shall be within  $\pm 10\%$  of their nominal values. Result comparisons will use truncated, whole integer values in units of mg/dL. These are inclusive ranges.
- d. All positive mixed volatile controls must meet these criteria for the batch to be acceptable for reporting of acetone, isopropanol and methanol. Failure to meet criteria for one mixed volatile compound does not invalidate the acceptability of another mixed volatile compound.
- e. If the positive mixed volatile control does not meet these criteria, the acceptability of the batch for ethanol reporting is not affected, provided the positive ethanol controls meet criteria in 10.11.3.2.
- f. Evaluation of ethanol performance in the mixed volatile positive control is not used in determining the acceptability of a batch for reporting of ethanol (see NOTE in 10.6.2.2).

#### 10.12 CRITERIA FOR CASE SAMPLE ACCEPTANCE

If the criteria for batch acceptance have been satisfied, the results of individual case samples are deemed suitable for reporting if the following criteria are met.

- 10.12.1 Chromatographic peaks for ethanol, acetone, isopropanol, methanol and n-propanol shall appear symmetrical.
- 10.12.2 The retention time for ethanol and n-propanol are  $\pm 2\%$  of those in ethanol calibrator 3. In mixed volatile data, the retention times for acetone, isopropanol, methanol and n-propanol are  $\pm 2\%$  of those in mixed volatile calibrator 3 (for quantitation) or the positive mixed volatile screening control (for initial identification). These are inclusive ranges.
- 10.12.3 Quantitative results for ethanol are reported from 0.01 - 0.40 g/100 mL for living subject samples and from 0.02 - 0.40 g/100 mL for postmortem samples. These are inclusive ranges.
- 10.12.4 Acetone, isopropanol and methanol are positively identified in a sample if the screening value is  $\geq 8$  mg/dL ( $\geq -20\%$  of 10 mg/dL target). Quantitative results for acetone, isopropanol and methanol are reported from 10 - 100 mg/dL. These are inclusive ranges.
- 10.12.5 When dilutions of case samples are tested, the quantitative result(s) before multiplication shall be within the reporting range of the test method.

#### 10.13 REPORTING

- 10.13.1 Blood ethanol results are reported according to the procedure found in the *Policy on Reporting of Blood Alcohol Results (P46-1)*.
- 10.13.2 Mixed Volatiles

- 10.13.2.1 Positive results for acetone, isopropanol and methanol are reported from the initial mixed volatile screen if values are  $\geq 8$  mg/dL, provided that the subsequent confirmation test (analyzed on a different GC column) yields a value  $\geq$  the LLOQ (10 mg/dL).
- 10.13.2.2 Quantitative results for acetone, isopropanol and methanol are reported as the whole integer, truncated result in units of mg/dL, to two significant figures (see NOTE in 10.7.1).
  - a. Example: an isopropanol value of 67.8 mg/dL is obtained.
    - The result is truncated to the whole integer value of 67 mg/dL (two significant figures) and reported.
- 10.13.2.3 Mixed volatile results may be reported as positive only, provided two tests have been performed, on two different GC columns, from individual samplings of the specimen.
- 10.13.3 When multiple dilutions are analyzed, the smallest dilution within the dynamic range is reported.
- 10.13.4 Liquor and cannabis board sample results are reported according to the *Policy on Testing and Reporting Results for Liquor and Cannabis Board samples (P48-3)*.
- 10.13.5 Quantitative results are reported for blood, plasma, serum, urine, vitreous and Liquor and Cannabis Board samples. Results for other specimen types are reported qualitatively (e.g., spleen squeeze, tissue homogenate).

#### 10.14 DOCUMENTATION AND REVIEW

- 10.14.1 In the event that a sequence is started on one day and completes after midnight, the date the sequence began will be the date of testing. Analysts will place their chromatograms, sequence tables and calibration tables and curves in a batch file.
- 10.14.2 The batch file will be forwarded to a reviewer for both a technical and administrative review. The reviewer will verify that the batch file contains all chromatograms, sequence tables and calibration tables and curves, all dates are correctly documented, the calibrator and control expiration dates have not been exceeded, individual chromatograms are initialed, all pages of the record are labeled with the batch number, and the calibrator and control values are within acceptable ranges. The reviewer will also verify that the batch meets the criteria for batch acceptance in 10.11 above.
- 10.14.3 The reviewer will sign and date the batch, indicating that the batch file is complete and the above procedures have been reviewed.
- 10.14.4 Upon completion of the technical and administrative review, the batch file is returned to the analyst.
- 10.14.5 The final batch file shall contain the calibration table and curves and all relevant sequence tables and chromatograms. Case sample chromatograms are filed in their respective case files.

#### 10.15 REFERENCES

1. Agilent (Hewlett Packard) 7694 Headspace Autosampler Operating and Service Manual.
2. Agilent (Hewlett Packard) 6890 Gas Chromatograph manual (Operating manual 1 and 2).
3. *Medicolegal Aspects of Alcohol*, fifth edition. Garriott JC, ed. 2008. Lawyers & Judges Publishing Company, Inc. Tucson, AZ.
4. *Disposition of Toxic Drugs and Chemicals in Man*, eighth edition. Baselt RC, ed. 2008. Biomedical Publications. Foster City, CA.

LIST OF CHANGES

Revision Date	Description	Page Number
08/20/12	Method approved by Washington State Toxicologist. See DRA dated 08/16/12. Method released for use in evidentiary testing on 08/20/12	All
10/09/12	Criteria for negative control acceptance changed to at or above 0.005 g/100 mL in section 10.10.3.1	5
09/19/14	Mixed volatile calibrators sourced as CRMs from approved supplier (10.6.2.2), calibrator levels changed to 10, 50 and 100 mg/dL (10.7). Changes made to criteria for batch (10.9, 10.10) and case sample acceptance (10.11), to reflect the new calibration levels. Added negative control criteria for mixed volatiles described in 10.10.3.1. Reporting of mixed volatiles detailed in 10.12. See DRA dated 9/12/14 for detailed changes.	2-7
07/07/15	Added note in 10.4 to describe testing for samples containing both sevoflurane and ethanol. Added section 10.8.5 to describe reinjection procedure. Modified criteria for source/preparation of mixed volatile positive control in 10.6.3.2 and 10.7.3.	1-5
11/17/15	Addition of 10.6.3.2 and changes to 10.7.1 describe use of a positive mixed volatile screening control for initial identification of mixed volatile compounds. Wording added to 10.7.3 for labeling/documentation of CTRL 4. Changes made to criteria for batch (10.9, 10.10) and case sample acceptance (10.11), to reflect initial screening for mixed volatile compounds. Reporting of mixed volatiles in 10.12.2 modified for qualitative reporting from initial screen and quantitative reporting from confirmation. Changed references from "Liquor Control Board" to "Liquor and Cannabis Board." Other minor edits throughout. See DRA dated 11/9/15.	All
7/24/17	Wording changed regarding dilutions in 10.4.3. Sequence amended in 10.7.1 to include use of a positive mixed volatile control (CTRL 4) at the end of confirmation batches that include quantitation of mixed volatiles (also added wording to NOTE in 10.7.1). Edited 10.8.5.3.b and 10.8.5.3.c to include specimen bracketing with CTRL 4. Reinjection reformatted as section 10.9, with numbering updated in subsequent sections. Changed acceptability criteria in 10.11.2.1.c, 10.11.2.2.c, 10.11.3.2.c and 10.11.3.3.c to specify that result comparisons use three decimal places for ethanol, one decimal place for mixed volatiles in calibrator 1, and the truncated, whole integer value for mixed volatiles calibrators 2 and 3 and positive control. Wording changed in 10.11.3.3.d to indicate all mixed volatile controls must pass for a target compound in order to report quantitative results for that compound. Added 10.13.5 to specify qualitative reporting of results for specific specimen types. Other minor edits throughout. See DRA dated 4/24/17.	All

